



# ISOLATION, IDENTIFICATION AND ANTAGONISTIC POTENTIAL OF *STREPTOMYCES* ANSCA22 FOR BIOCONTROL OF *RHIZOCTONIA BATATICOLA* INFECTION OF SOYBEAN

Amarja S. Khendkar\* and Aarti R. Deshpande

Department of Microbiology, Shankarlal Khandelwal College, Akola (Maharashtra), India.

## Abstract

*Streptomyces* ANSCa22 isolated from rhizospheric soil of *Capsicum annuum* demonstrated antifungal activity against the pathogen *Rhizoctonia bataticola* by dual culture technique. *Streptomyces* ANSCa22 treatment on *Rhizoctonia bataticola* inoculated Soybean seeds could control *Rhizoctonia bataticola* infection of Soybean in pot study which was evident from the significant increase in the values of average shoot height, pod number, seed weight and plant dry weight after the treatment. *Streptomyces* ANSCa22 treatment of normal soybean seeds without any pathogen inoculation also resulted in positive effect on three out of four tested plant growth parameters as compared to control. Microscopic examination of *Streptomyces* ANSCa22 revealed typical features of *Streptomyces*. Partial sequencing of rRNA gene identified ANSCa22 as *Streptomyces rochei*. This strain was compatible with *Bradyrhizobium japonicum*, exhibited tolerance to calcium carbonate and calcium sulphate upto 5% concentration and indicted varying degree of growth at 7 to 10 pH range. *Streptomyces rochei* ANSCa22 was positive for amylase enzyme.

**Key words :** Antagonistic *Streptomyces*; *Rhizoctonia bataticola*; Biocontrol.

## Introduction

Management of *R. bataticola* infection of Soybean is of major concern in Akola district owing to the prevalence of this fungal phytopathogen. Root rot, Stem rot, Charcoal rot and damping-off are some of the diseases caused by this pathogen resulting in yield losses (Yang and Navi, 2005, Mengistu *et al.*, 2011). In major soybean growing states of India, incidence of *Rhizoctonia* root rot has been reported in the range of 3.29% to 40.33% and in Maharashtra it was in the range of 17.49% to 37.44% during 2012 to 2013. (Belkar. and Gade, 2016).

Chemical fungicides have been commonly employed to control these diseases (Divya and Sudini 2013 ). However chemical control has always been associated with adverse effects of chemical fungicides on environment and agro- ecosystem (Yang *et al.*, 2011). During past few decades the efforts have been focused on exploring eco-friendly and safe biocontrol agents for management of plant diseases (Ratul and Singh, 2018).

Importance of *Streptomyces sp.* for biocontrol of phytopathogens and plant growth promotion has been well established by many researchers. (Gopalakrishnan *et al.*, 2011 b; Viswanathan *et al.*, 2015; Ahmed 2017).

*Streptomyces* Strains Di944 (Siva, 1999) and *Streptomyces lydicus* 108 (Yuan and Crawford, 1995) have been patented in Landon and United state respectively. Focused studies at International Crops Research Institute for the Semi-Arid Tropics (ICRISAT) of Andhra Pradesh, India ( Gopalkrishana *et al.*, 2012) have resulted in promising outputs with respect to agricultural applications of *Streptomyces* spp.

The success of the applications of any microbial agro-input is directly related with the acquaintance of the strain with the physico-chemical and the climatic conditions prevailing in the agro-ecosystem of the region. The present investigations deal with isolation, identification and evaluation of antagonistic and plant growth promoting potential of *Streptomyces* ANSCa22, an indigenous strain of Akola District, Maharashtra, India.

## Materials and Methods

### Isolation of *Streptomyces*

Rhizospheric soil sample was collected from *Capsicum annuum* from a field of Kanheri sarap village of Akola district. Streptomycetes were isolated by serial dilution method (Usha and Selvam 2011) on Starch casein agar (SCA). Compact, hard, powdery, slow growing colonies were selected. Pure cultures were maintained in glycerol stock.

### Isolation of *R. bataticola*

Soybean plant samples were collected from the fields of North, South, East and West side villages of Akola district. Diseased plants looking different from the normal healthy plants with respect to general appearance, height and colour of leaves were collected in different carry-bags. Surface sterilised small pieces of roots or stems were placed on PDA and incubated for 7 days at 27°C. Isolated fungal plant pathogens were identified on the basis of colony characteristics and microscopic examination described by (Veena *et al.*, 2014).

### Antifungal activity by dual culture technique

*R. bataticola* was inoculated on SCA by spread plate technique. *Streptomyces* isolates were spot inoculated on the same plate. After 7 days incubation colonies showing zone of inhibition against *R. bataticola* were selected.

Antifungal activity of *Streptomyces* ANSCa22 against *R. bataticola* was studied further by inoculating the two cultures on the same SCA plate opposite to each other (Gopalkrishana *et al.*, 2012).

### Microscopic examination of selected isolate

Cover slip method was used (Williams *et al.*, 1983) in which half of sterilized cover slip was inserted in the starch casein agar plate. Culture of isolate ANSCa22 was inoculated (straight line) near cover slip on agar. After incubation period cover slip was removed and observed under microscope. The culture was identified on the basis of arrangement of spores, according to Madigan *et al.*, (1987).

### Test for Pathogenicity of *R. bataticola*

Inoculum of *R. bataticola* was mixed into sterilized soil at 4% by weight of soil (Lukade, 1992) and was filled in sterilized pots. Surface sterilized Soybean seeds were used for sowing. Soil without inoculum of pathogen was used as control. Readings were noted on 15th day after sowing regarding germination % and number of survived and infected plants. Percent disease incidence was calculated (Srinivas, 2016). *R. bataticola* isolate (R1)

demonstrated 100% disease incidence.

### Pot study

Fresh biomass (20%) of *Streptomyces* ANSCa22 was mixed with talcum powder. Fresh inoculum of *R. bataticola* (R1) was prepared in potato dextrose broth. Surface sterilized seeds were soaked in broth of *R. bataticola* (R1) for 30 minutes and then the seeds were subjected to different treatments.

These treatments included R1 inoculation followed by *Streptomyces* ANSCa22 treatment (100mg/g seeds) (T<sub>1</sub>) and Carbendazim treatment (0.2mg/g of seeds) (T<sub>3</sub>). Other pots of the sets were R1 inoculated seeds without any treatment (T<sub>4</sub>), only *Streptomyces* ANSCa22 treated (100mg/g) seeds without R1 inoculation (T<sub>2</sub>) and control (C1) *i.e.* seeds without any treatment. (Sabaratnam, 1999). The effects on seed germination and plant growth parameters were monitored for three months using sterilized pots and soil.

### Statistical analysis

Statistical analysis was carried out by using Graph pad prism software version 5. ANOVA and T test were performed (Otto-Hanson *et al.*, 2012).

### Stress tolerance for salt concentration and pH

*Streptomyces* ANSCa22 was grown on Starch casein agar media with different concentrations of NaCl (2.5%, 5%, 7.5% and 10%), CaCO<sub>3</sub> (2.5% and 5%), CaSO<sub>4</sub> (2.5% and 5%) and CaCl<sub>2</sub> (2.5% and 5%), and on different pH (7, 8, 9, 10) (Carla *et al.*, 2008, Sunita *et al.*, 2015).

### Cellulose degradation

*Streptomyces* ANSCa22 isolate was spot inoculated on Czepak mineral salt medium and incubated for 2-3 days at 28°C. After incubation period plate was flooded with 1% Congo red solution and excess dye was drained after 5 minutes. Solution of 1M NaCl was then added repeatedly until colour of Congo red solution disappeared. Clear zone around the colony indicated degradation of cellulose (Carder 1986; Sharma *et al.*, 2001).

### Chitin degradation

*Streptomyces* ANSCa22 isolate was spot inoculated on colloidal chitin agar plate (swollen chitin 2g/lit, ammonium sulphate 0.05g/lit., agar 1.8g/lit. and pH 7) and incubated up to 5 days. Chitinase activity was identified by clear zone around the colony (Palaniyandi *et al.*, 2013).

### Hydrolysis of casein

*Streptomyces* ANSCa22 isolate was spot inoculated on casein agar plate (10g/lit. casein, 10g/lit., glucose, 1.5g/lit., K<sub>2</sub>HPO<sub>4</sub> and 15g/lit. agar) (Palaniyandi *et al.*, 2013)

and incubated at 28°C for 5 days, Casein hydrolysis was identified by clear zone around the colony.

### Hydrolysis of gelatin

*Streptomyces* ANSCa22 isolate was spot inoculated on nutrient agar containing 15% gelatin and incubated for 3 to 4 days. After incubation plate was flooded with 1% HgCl<sub>2</sub>. Clear zone around the colony indicated the hydrolysis of gelatin (Stefka *et al.*, 2004).

### Hydrolysis of starch

*Streptomyces* ANSCa22 isolate was spot inoculated on starch agar plate and incubated at 28°C for 3-5 days. After incubation period plate was flooded with 1% iodine. Clear zone around the colony indicated the amylase activity (Shaw *et al.*, 1984; Santos *et al.*, 2003).

### Compatibility with *Bradyrhizobium japonicum*

*Bradyrhizobium japonicum* was inoculated on SCA by spread plate technique. *Streptomyces* ANSCa22 was spot inoculated on the same plate. After 9 days incubation growth of both microorganisms without zone of inhibition against *Bradyrhizobium japonicum* indicated compatibility of *Streptomyces* ANSCa22 with *Bradyrhizobium japonicum*.

### Identification of Selected *Streptomyces*

Molecular phylogeny of *Streptomyces* ANSCa22 was determined by amplifying and sequencing genomic 16s rRNA region using 8F and 1541R primers. Sequencing facility was availed from Yaazh Xenomics Chennai. The results obtained in this study were used for identification of isolates.

## Results and Discussion

Total 12 cultures were isolated from rhizospheric soil of Capsicum (*Capsicum annum*) viz., ANSCa22 to ANSCa33. All isolates exhibited typical characteristics of *Streptomyces* colony (compact, hard, powdery, slow growing colony). *R. bataticola* was isolated from infected plant sample. Brown to black colour mycelial growth of *R. bataticola* (R1) was observed on agar. In microscopic examination, dark

sclerotia with round to irregular shape and right angle branch to parent hyphae were observed (Fig. 1).

Colony of *Streptomyces* ANSCa22 (Fig. 2) had white colour arial mass and yellow colour reverse side pigment. No melonoid pigment production was seen. Spiral and hook like) spore arrangement was observed in microscopic examination of *Streptomyces* ANSCa22 (Fig. 3). Screening of twelve *Streptomyces* isolates for antifungal activity against R1 by dual cultures technique revealed the presence of antagonistic activity in ANSCa22 (Fig. 4). None of the 11 other isolates were antagonistic for the pathogen R1.

*Streptomyces* ANSCa22 was studied for the antagonistic potential against the pathogen R1 and also for plant growth promoting potential for soybean in pots. Averages of shoot height, number of pods, seed weight and plant dry weight of the pathogen R1 inoculated seeds were studied after different treatments. *Streptomyces* ANSCa22 treatment (T<sub>1</sub>) could effectively control *R. bataticola* infection as evident from improvement in all the growth parameters after the treatment (Table 1). Efficacy of this biological treatment was appreciably better than the chemical treatment with carbendazim (T<sub>3</sub>)

**Table 1:** Effect of different treatments on growth parameters of soybean.

Treatments	Average of shoot height (cm)	Average of pod no.	Average of seed weight (gm)	Average of plant dry weight
Control (C1)	36.74	8.800	0.7316	12.70
<i>R. bataticola</i> + <i>S. ANSCa22</i> (T <sub>1</sub> )	36.20	13.00	0.970	16.24
<i>Streptomyces</i> ANSCa22 (T <sub>2</sub> )	40.20	14.40	0.981	17.00
<i>R. bataticola</i> + Carbendazim (T <sub>3</sub> )	15.54	4.400	0.5208	6.640
<i>R. bataticola</i> (T <sub>4</sub> )	1.480	0.200	0.0320	1.460

**Table 2:** P values of T test and ANNOVA for results of pot study.

S.No.	Parameters checked	T test (P value)		ANOVA (P value) (Between 5 groups)
		T1vs T4	T1vs C1	
1	Plant height	0.0097	0.7518	0.0012
2	No. of Pods	0.0097	0.0119	0.0003
3	Seeds weight	0.0097	0.0079	0.0002
4	Plant dry weight	0.0097	0.0079	0.0003

Note: T<sub>1</sub>- *R. bataticola* and *Streptomyces* ANSCa22 treated seeds, T<sub>4</sub> - Only *R. bataticola* treated seeds and C1- Control.

**Table 3:** Effect of different salt concentrations on the growth of *Streptomyces* ANSCa22 isolate.

S.No.	Isolates	CaCo3		CaSO4		CaCl2		NaCl	
		2.5%	5%	2.5%	5%	2.5%	5%	2.5%	5%
1	ANSCA22	++++	++	+++	++	+	-	+	+

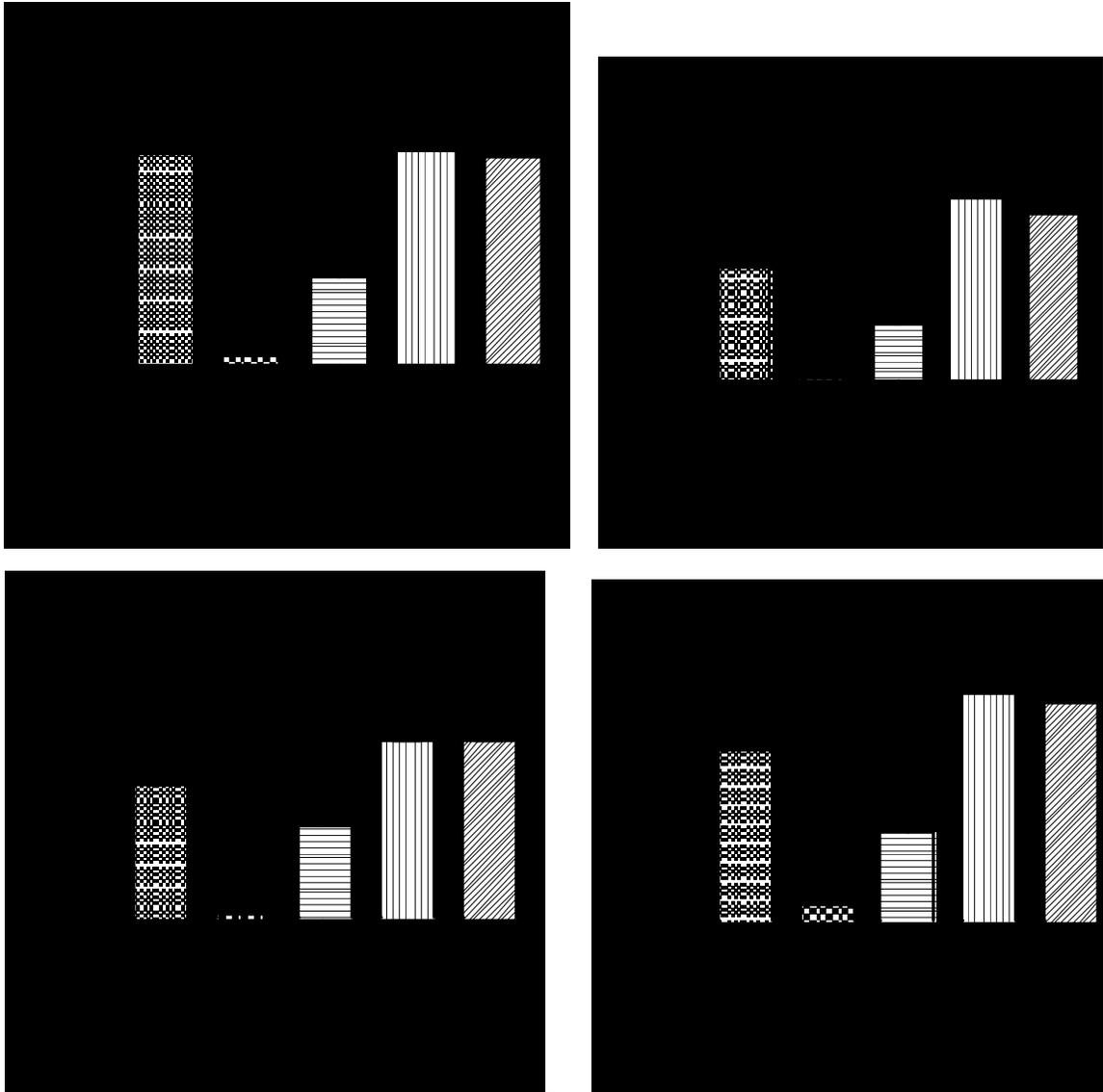
Poor growth +, moderate growth ++, good growth +++, luxuriant growth +++++, No growth -



**Fig. 1:** Microscopic examination of **Fig. 2:** *Streptomyces rochei* ANSCa22 *R. bataticola*.



**Fig. 3:** Microscopic examination of **Fig. 4:** Dual plate technique. *Streptomyces rochei* ANSCa22



**Note:** C1-Control, T4 (R1)-*R. bataticola*, T5-*R. bataticola* and Carbendazim treated seeds, Only ANSCa22, *R. bataticola* and *Streptomyces* ANSCa22 treated seeds.

**Fig. 5:** Effect of different treatments on growth parameter of Soybean.

( Fig. 5). Average Shoot height, pod number, seed weight and plant dry weight increased after *Streptomyces* ANSCa22 treatment by 36.1cm, 12, 0.947g and 15.74g respectively and after chemical treatment the increase in the four above parameters was 13.66cm, 4.2, 0.488g

and 5.18g respectively.

*Streptomyces* ANSCa22 treatment (T<sub>2</sub>) of normal soybean seeds (without pathogen inoculation) also resulted in remarkable improvement in three out of four tested growth parameters as compared to control (C1)



**Fig. 6:** Compatibility with *B. japonicum*  
**Fig. 7:** Starch hydrolysis.

(Table 1, Fig. 5 ). This clearly indicated the presence of plant growth promoting potential for soybean in *Streptomyces* ANSCa22.

Significance of the observations was analyzed by T test and ANOVA. P values obtained were in the range of 0.0079 to 0.0119 in T test and 0.0002 to 0.0012 in ANOVA. These readings supported the significance of plant growth promoting potential and antagonistic bioactivities of *Streptomyces* ANSCa22 culture (Fig. 5, Table 2).

The presence of two favourable attributes of antagonism and plant growth promotion in *Streptomyces* ANSCa22 is very beneficial for the field application of the culture. Moreover, *Streptomyces* ANSCa22 appeared to be compatible with *Bradyrhizobium japonicum*. ( Fig.6) Many earlier studies have reported the presence of both the antagonistic and plant growth promoting activities together in one culture of *Streptomyces* species (Sabrtnam, 1999, Gopalakrishnan *et al.*, 2011b).

*Streptomyces* ANSCa22 could grow at varying degree at 2.5% and 5% concentrations of NaCl, CaCO<sub>3</sub> and CaSO<sub>4</sub> table 3 but not at 5% CaCl<sub>2</sub> concentration. *Streptomyces* ANSCa22 was also found to grow on pH 7 to 10 (luxuriant growth to poor growth). Thus its application potential appears to be suitable for neutral to alkaline and non saline to slightly saline soils.

*Streptomyces* ANSCa22 was observed to hydrolyze starch (Fig. 7) but negative results were obtained about chitin, cellulose, gelatine and casein degradation.

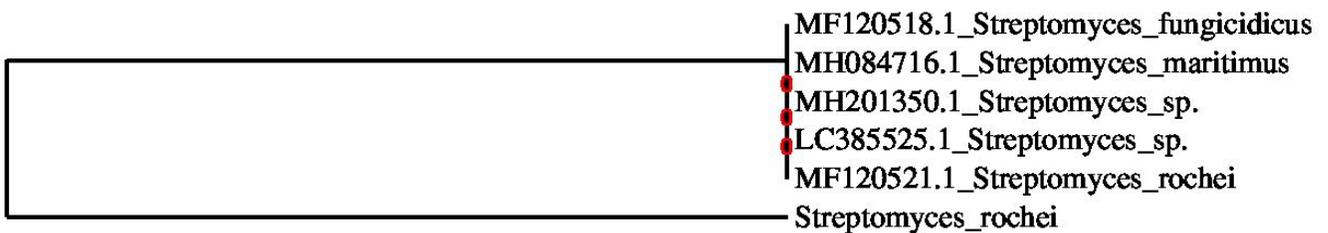
*Streptomyces* ANSCa22 isolate was identified by

16s rRNA gene sequencing. Polymerase chain reaction was carried out using 8F and 1541R primer. The 16s rRNA sequence was blast using NCBI blast similarity search tool. It was identified as *Streptomyces rochei*.

The nucleotide sequence of this organism is

>CONTIG STREPTO

```
GAGTTGCAGACCCCAATCCGAACTGAGACCGG
CTTTTTGAGATTCGCTCCACCTCGCGGTATCGCA
GCTCATTGTACCGGCCATTGTAGCACGTGTGCA
GCCAAGACATAAGGGGCATGATGACTTGAC
GTTCGTCACCTTCCCTCCGAGTTGACCCCGGC
GGTCTCCCGTGAGTCCCCAGCACCACAAGGGCC
TGCTGGCAACACGGGACAAGGGTTGCGCTCGTTGCGG
GACTTAACCCAACATCTCACGACACGAGCTGA
CGACAGCCATGCACCACCTGTACACCGACCACAAGG
GGGACCCCTGTCTCCAGGGTTTTCCGGTGT
TATGTCAGCCTTGTTAAGGTTCTT
CGCGTTGCGTTCGAATTAAG
CCACATGCTCCGCCGCTTGTGC GGGCCCCC
GTCAATTCCTTTGAGTTTTAGCCTTGGCGCCGTACTCC
CAGGCGGGGCACTTAATGCGTTAGCTG
CGGCACGGACAACGTTGGAAATG
TTGCCACACCTAGTGCCACCGTTTACGGCG
TGGA CTACCAGGGTATCTAATCCTGTTC
GCTCCCCACGCTTTCGCTCCTCAGCGTCAGTA
TCGGCCCAGAGATCCGCTTCGCCACC
GGTGTTCCTCCTGATATCTGCGCATTTCACCGCTA
CACCAGGAATTCGATCTCCCCTACCGA ACTCT
AGCCTGCCCCGATCGACTGCAGACCCGGGGTTAA
GCCCCGGGCTTTCACAACCGACGTGACAAGCCGC
CTACGAGCTCTTTACGCCAATAATTCGG
GACAACGCTTGCGCCCTACGTATTACCGCGGCT
GCTGGCACGTAGTTAGCCGGCGCTTCTTC
TGCAGGTACCGTCACTTTCGCTTCTCCCTGCTGAAA
GAGGTTTACAACCCGAAGGCCGTCATC
CCTCACGCGGCGTCGCTGCATCAGGCTTTCGCCCA
TTGTGCAATATCCCCACTGCTGCCTCCCGTAG
GAGTCTGGGCCGTGTCTCAGTCCCAGTGTGGCCG
GTCGCCCTCTCAGGCCGGCTACCCGTCGTCGCCTTGGT
GAGCCGTTACCTACCAACTAGCTGATAGGCCGC
```



GGGCTCATCCTGCACCGCCGGAGCTTTTCGAACC  
TCGCAGATGCCTGCGAGGATCAGTATCCGGTATTA  
GACCCCGTTTCCAGGGCTTGTCCCAGAGTGCAGGG  
CAGATTGCCACGTGTTACTACCCGTTTCGCCACT  
AA

Sequence of *Streptomyces rochei* ANSCa22 deposited under the GenBank accession number is SUB4723523 Strep MK116539.

### Conclusion

*S. rochei* ANSCa22 appeared to possess antagonistic as well as plant growth promoting potentials with respect to *R. bataticola* infection and plant growth of soybean.. Its biocontrol treatment was more effective as compared to Carbendazim treatment for all the growth parameters in pot trails. Hence this culture needs to be explored with intensive studies to evaluate its application potential in fields.

### Acknowledgements

Authors are thankful to Shankarlal Khadelwal College, Akola, for providing required facilities to carry out this study and Dr B.T. Raut, Ex Head of Plant Pathology Dept. Dr. Panjabrao Krushi Vidyapeetha, Akola.

### References

- Abd-Elsalam, K.A. (2010). Genetical and biological control of cotton ashy stem caused by *Macrophomina phaseolina* in outdoor pot experiment. *Saudi Journal of Biological Sciences*, **17**: 147–152.
- Ahmed, I.S. Ahmed (2017). Efficacy of two potent soilborne *Streptomyces* spp. In Controlling wilt and root rot diseases of tomato (*lycopersicon esculentum* mill) under greenhouse conditions. *Int. J. Pharm. Bio. Sci.*, **8(3)**: 394-403.
- Belkar Y.K. and R.M. Gade (2016). Survey for Incidence of *Rhizoctonia* Root Rot in Major Soybean Growing States. *Advances in Life Sciences*, **5(10)**.
- Bonaldi, M., X. Chen, A. Kunova, C. Pizzatti, M. Saracchi and P. Cortesi (2015). Colonization of lettuce rhizosphere and roots by tagged *Streptomyces*. *Front. Microbiol.*, 1-6.
- Carder, J.H. (1986). Detection and quantitation of cellulase by Congo red staining of substrates in a up-plate diffusion assay. *Anal. Biochem.*, **153(1)**: 75-79.
- Carla da Silva Sousa, Ana Cristina Fermino Soares, Marlon da Silva Garrido (2008). Characterization of Streptomycetes with potential to promote plant growth and biocontrol. *Sci. Agric. (Piracicaba, Braz.)*, **65(1)**: 50-55.
- Elango, V., M.K. Kolandasamy, P. Ponmurugan and S. Marimuthu (2015). "Evaluation of *Streptomyces* spp. for effective management of *Poria hypolateritia* causing red root-rot disease in tea plants". *Biological Control*, **89**: 75–83.
- Divya Rani, V. and H. Sudini (2013). Management of soilborne diseases in crop plants: An overview. *International Journal of Plant, Animal and Environmental Sciences*.
- Gopalakrishnan, S., B.K. Kiran, P. Humayun, M.S. Vidya, K. Deepthi and O. Rupela (2011b). Biocontrol of charcoal-rot of sorghum by Actinobacteria isolated from herbal vermicompost. *Afr. J. Biotechnol.*, **10**:18142-18152.
- Gopalakrishnan, S., P. Humayun, S. Vadlamudi, R. R.K. Vijayabharathi, Bhimineni and O. Rupela (2012). "Plant growth-promoting traits of *Streptomyces* with biocontrol potential isolated from herbal vermicompost". *Biocontrol Sci. Technol.*, **22**: 1199-1210.
- Houssam, O.M. (2009). An Antifungal Agent Produced by *Streptomyces olivaceiscleroticus*, AZ-SH514. *World Applied Sciences Journal*, **6(11)**: 1495-1505.
- Jeffrey Lim, S.H., M.S. Umi Kalsom, A.R. Nor Aini, S. Khozirah and H. Halizah (2015). *Streptomyces* ambofaciens S2 - A Potential Biological Control Agent for *Colletotrichum gloeosporioides* the Causal Agent for Anthracnose in Red Chilli Fruits. *J. Plant Pathology Microbiology*, **S:1**.
- Khendkar, A.S. and A.R. Deshpande (2018). Isolation and screening of *Streptomyces* for biocontrol potential against *Rhizoctonia bataticola* infection of soybean. *Indian J. of Appl. Microbio.*, **21(1)**: 78-86.
- Konde, S.W. and B.T. Raut (2008). Management of Root/Collar Rot Disease in Soybean. *Journal of Plant Disease Sciences*, **3**: 81-83.
- Kunova, A., M. Bonaldi, M. Saracchi, C. Pizzatti, X. Chen and P. Cortesi (2016). Selection of *Streptomyces* against soil borne fungal pathogens by a standardized dual culture assay and evaluation of their effects on seed germination and plant growth. *BMC Microbiology*, **272**: 1-11.
- Morton, D.T. and N.H. Stroube (1955). Antagonistic and stimulatory effect of microorganism upon *Sclerotium rolfsii*. *Phytopathology*, **45**: 419–420.
- Otto-Hanson, L.K., Z. Grabau, C. Rosen, C.E. Salomon and L.L. Kinkel (2012). Pathogen Variation and Urea Influence Selection and Success of *Streptomyces* Mixtures in Biological Control. *Phytopathology*, **103(1)**: 35-42.
- Palaniyandi, S.A., H.Y. Seung, B. Karthiyaini and S. Joo-Won (2013). Genetic and functional characterization of culturable plant-beneficial actinobacteria associated with yam rhizosphere. *J. Basic Microbiol.*, 1 - 11.
- Ratul Ram, M. and B. Singh Harikesh (2018). Biocontrol Technology: Eco-friendly approaches for sustainable agriculture. *Omics Technologies and Bio engineering*, **2**: 177-190.
- Santos, E.O. and M.L.L. Martins (2003). Effect of the medium composition on formation of amylase by *Bacillus* sp. *Braz. Arch. Biol. Technol.*, **46(1)**: 129-134.

- Sabaratnam, S. (1999). Biological control of *rhizoctonia* damping-off of tomato with a rhizospheric Actinomycetes. Thesis, Faculty of Graduate Studies University of Western Ontario London, Ontario.
- Sakure, S., A. Limbore, M. Zalake and S. Jaigude (2015). Isolation and characterization of *Actinomycetes* from rhizosphere Soil of different Plants for anti-phytopathogenic activity and stress tolerance. *Int. J. Curr. Microbiol. App. Sci.*, **2**: 379-387.
- Skidmore, A.M. and C.H. Dickinson (1976). Colony interactions and hyphal interference between *Septoria Nodorum* and phylloplane fungi. *Trans. Brit. Mycol. Soc.*, **66**: 57-64.
- Sharma, R., Y. Chisti and U.C. Banerjee (2001). Production, purification, characterization, and applications of lipases. *Biotechnol. Adv.*, **19**: 627-662.
- Shaw, F.J. and T.M. Ou-Lee (1984). Studies on the amylase from the germinated rice seeds. *Bot. Bull. Acad. Sin.*, **23**: 41-46.
- Stefka, A.N., T. Nikoleta and Y. Ljubomira (2004). Taxonomy of *Streptomyces* sp. strain 3B. *J. Cult Collect.*, **4**: 36-42.
- Xiao, K.L., L.L. Kinkel and A.D. Samac (2002). Biological control of Phytophthora root rot on alfalfa and soybean with *Sterptomyces*. *Biol Control.*, **23** : 285–295.
- Yang, C., C. Hamel, V. Vujanovic and Y. Gan (2011). Fungicide: Modes of Action and Possible Impact on Non target Microorganisms. *ISRNEcology*. doi:10.5402/2011/130289
- Yuan, W.M. and D.L. Crawford (1995). Characterization of *Streptomyces lydicus* WYEC108 as a potential biocontrol agent against fungal root and seed rots. *Applied and Environmental Microbiology*, **61(8)**: 3119–3128.